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Separation of Dissolved Iron from the Aqueous System with Excess Ligand

Hiroshi Hasegawa,*1 Ismail M. M. Rahman,*1, 2 Sanae Kinoshita,1 Teruya Maki,1 and Yoshiaki Furusho3

1Graduate School of Natural Science and Technology, Kanazawa University, Kakuma, Kanazawa 920-1192, Japan

2Department of Chemistry, University of Chittagong, Chittagong 4331, Bangladesh

3GL Sciences, Inc., Nishishinjuku 6-22-1, Shinjuku, Tokyo 163-1130, Japan

*Author(s) for correspondence.
E-mail: hhiroshi@t.kanazawa-u.ac.jp (H. Hasegawa), I.M.M.Rahman@gmail.com (I.M.M. Rahman)
Tel/ Fax: +81-76-234-4792
Abstract

A new technique for the separation and preconcentration of dissolved Fe(III) from the ligand-rich aqueous system is proposed. A solid phase extraction (SPE) system with an immobilized macrocyclic material, commonly known as molecular recognition technology (MRT) gel and available commercially, was used. Synthetic Fe(III) solution in aqueous matrices spiked with a 100-fold concentration of EDTA was used. Dissolved iron that was ‘captured’ by the MRT gel was eluted using hydrochloric acid and subsequently determined by graphite furnace atomic absorption spectrometry. The effect of different variables, such as pH, reagent concentration, flow rate and interfering ions, on the recovery of analyte was investigated. Quantitative maximum separation (~100%) of the dissolved Fe(III) from synthetic aqueous solutions at a natural pH range was observed at a flow rate of 0.2 mL min⁻¹. The extraction efficiency of the MRT gel is largely unaltered by the coexisting ions commonly found in natural water. When compared with different SPE materials, the separation performance of MRT gel is also much higher.

Keywords

Dissolved iron; preconcentration; excess ligand in aqueous solution; Molecular Recognition Technology; atomic absorption spectrometry
1.0 Introduction

The accurate determination of iron in dissolved and particulate forms or size fractions is necessary for describing the iron chemistry within a given body of water (Bruland and Rue, 2001). The iron content in an aqueous system is important for environmental protection, hydrogeology and some chemical processes, such as photosynthesis and phytoplankton production in the open oceans (Riley and Chester, 1971; Martin and Fitzwater, 1988; Martin, 1990). Recent studies suggest that Fe(III) solubility in oceanic water is controlled by complexation with natural organic ligands (Kuma et al., 1996; Waite, 2001; Liu and Millero, 2002), and these complexation equilibria subsequently regulate the concentrations of dissolved iron in the oceanic system (Johnson et al., 1997; Kuma et al., 2003). Thus, the majority of dissolved iron in the oceans appears to be chelated (as FeL₄) with organic ligands (L₄) (Wu and Luther, 1995; Rue and Bruland, 1997; Boye et al., 2001).

Sensitive analytical methods, such as graphite furnace atomic absorption spectrometry (GF-AAS), have led the way in providing reliable profiles for iron in oceanic waters over the past three decades (Johnson et al., 1997; Bruland and Rue, 2001). However, precise determination of iron is very difficult due to the risk of sample contamination (Blain and Treguer, 1995; de Jong et al., 2008). Reports have shown that there are discrepancies between the estimated concentrations of dominant hydroxo-complex species of Fe(III), such as Fe(OH)₂⁺, Fe(OH)₃⁰ and Fe(OH)₄⁻, and values (~0.1–10 nM) for the thermodynamic solubility of Fe(III) hydroxide in seawater (Byrne and Kester, 1976; Kuma et al., 1996; Liu and Millero, 2002). These low concentrations also make it difficult to measure iron in ocean waters (de Jong et al., 2008). For reliable analysis, method development is important to overcome limitations caused by the high salt matrix of oceanic water samples that could cause interferences during measurements. One of
the popular analysis methods is to separate and preconcentrate the desired metal ion(s) out of the sample matrix for precise determination (Hosten and Welz, 1999). Various methods, such as co-precipitation, liquid-liquid extraction, solid-liquid extraction, and ion exchange resins, have been developed for the separation of trace metals from natural samples (Filik et al., 1997; Pourreza and Mousavi, 2004; Ghaedi et al., 2005; Ghaedi et al., 2007; Ghaedi et al., 2008). However, the solid phase extraction (SPE) approach has gained rapid acceptance since the 1980s because it is one of the fastest, most economical and cleanest methods for separating and concentrating trace metals from aqueous samples (Hosten and Welz, 1999; Firdaus et al., 2007).

In SPE, appropriate materials are immobilized onto support matrices to prepare solid phases with a ‘capturing capability’ for separation and preconcentration (Masi and Olsina, 1993). Some of the sorbent materials have the ability to interact with various metal ions while others are fairly specific for a particular ion within complex matrices (Carbonell et al., 1992; Nickson et al., 1995; Ghaedi et al., 2006; Ghaedi et al., 2008; Ghaedi et al., 2009). One group of SPE materials includes those with macrocyclic ligands, such as crown ethers, immobilized on a silica or polymer support; this type of SPE has been reportedly used for ion-selective separation and preconcentration of metal ions, and the technique is commonly known as molecular recognition technology (MRT) (Izatt et al., 1995; Hasegawa et al., 2010; Rahman et al., 2010). Reports on the separation/preconcentration of dissolved iron using SPE from an aqueous system in which Fe-organic ligand complexation is prevalent is still very limited (Wells and Bruland, 1998). The extraction efficiency of SPE materials significantly decreases in aqueous systems containing excess ligand because ligands often compete with SPE materials for metal ions. This limitation can be minimized with the MRT-SPE systems, which can provide non-destructive separation of metal ions from ligand-rich aqueous systems (Hasegawa et al., 2010).
In this work, we propose a simple method using a MRT-SPE system for the separation of dissolved iron from aqueous system of Fe-ligand complexes containing an excess amount of ligand in solution. The SPE sorbent is a proprietary polymeric organic material comprised of ion-selective sequestering agents based on molecular recognition and macrocyclic chemistry. Synthetic, ligand-rich aqueous Fe-ligand matrices were used as the samples. Hydrochloric acid was used as the eluent with subsequent determination of iron using GF-AAS.

2.0 Experimental

2.1 Materials

2.1.1 Reagents

Analytical grade commercial products were used as received. Stock solutions of Fe(III) and Fe(II) were prepared by dissolving FeCl₃·6H₂O and FeCl₂·4H₂O (Wako Pure Chemical Industries, Japan), respectively, in 1 M HCl. A Perkin-Elmer (Norwalk, CT, USA) Model ICP Optima 3000 inductively coupled plasma emission spectrometer was used to standardize the iron solutions.

Nitrilotriacetic acid (NTA), O,O’-bis(2-aminoethyl)ethyleneglycol-N,N’,N’’-tetraacetic acid (GEDTA), ethylenediamine-N,N,N’,N’’-tetraacetic acid (EDTA) and diethylenetriamine-N,N,N’,N’’-pentaacetic acid (DTPA) were purchased from Dojindo Laboratories, Japan; desferrioxamine B (DFB) was supplied by Sigma-Aldrich, St. Louis, MO. Each of these reagents was dissolved in 0.1 M sodium hydroxide to prepare stock solutions of the chelating ligands.

Working standards of metal and ligand solutions in the µM to mM range were prepared by dilution on a weight basis. The experimental pH range was 4 to 8, and solution pH was adjusted.
using either 1 M HCl or 1 M NaOH. The pH was maintained using the following buffer solutions: 0.1 M CH₃COONa/CH₃COOH (pH 4–5) and 0.1 M NaH₂PO₄/Na₂HPO₄ (pH 6–8).

Aqueous solutions of 10 mM chelating ligands in the appropriate buffer were spiked with 0.1 mM Fe(III) or Fe(II) in 1.0 M HCl, and allowed to stand for 20 min, and these solutions were then added to the sample solutions. For Fe(II) sample solutions, 0.2% hydroxyl ammonium chloride was added to prevent oxidation.

To study the effect of coexisting ions, NaCl, KCl, CaCl₂ and MgCl₂ were used as sources of cations, and NaCl, NaNO₃, CH₃COONa, Na₃PO₄, Na₂SO₄, NaClO₄ were used as sources of anions; all salts were purchased from Nacali Tesque, Japan. Working solutions at a concentration of 10 mM were prepared in a H₂O matrix, and the pH was adjusted to 8.0. The final solutions were allowed to equilibrate for 24 h before analysis. Considering the competitive behavior of Ca²⁺ and Mg²⁺ ions in seawater in terms of ligand capturing, the metal-to-ligand ratio was maintained at 1:100.

Deionized water obtained from a Barnstead 4 Housing E-Pure system was used to prepare all solutions and is hereafter referred to as EPW.

2.1.2 SPE materials

Different SPE materials, as listed in Table 1, were used. MRT gel and AnaLig TE-01 were purchased from GL Sciences, Japan. Other SPE materials were purchased from Bio-Rad Laboratories (Chelex-100), GL Sciences (MetaSEP ME-1, ME-2 and ME-3) and Hitachi High-Technologies Corporation (NOBIAS Chelate PA-1, PB-1, Ion SC-1 and SA-1).

2.2 Cleaning

Low-density polyethylene and laboratory equipment (Nalge, USA) were used to store the solutions and to hold solutions during the experiments. Before use, bottles and laboratory
equipment were soaked in an alkaline detergent (Scat 20X-PF, Nacali Tesque, Japan) overnight and rinsed with EPW; they were then soaked in 4 M HCl overnight and rinsed with EPW. PerFluoroAlkoxy tubes and micropipette tips (Nichiryo, Japan) were cleaned according to the procedure described by Sohrin et al. (1998).

2.3 Column separation procedure

2.3.1 Column cleaning and conditioning

SPE materials packed in 3-mL columns were used in this experiment. Column cleaning was conducted with HNO₃ (8 mL) and EPW (6 mL). The appropriate buffer solution, consisting of 5 mL of 0.1 M CH₃COONa/CH₃COOH (pH 4–5) or 5 mL of 0.1 M NaH₂PO₄/Na₂HPO₄ (pH 6–8), was allowed to flow through the column to ensure the desired pH conditions (4–8).

2.3.2 Treatment of samples

Sample solution (5 mL) with ligand (10 mM) and spiked with Fe(III) or Fe(II) (0.1 mM), which was pH adjusted with 0.1 M solution of pH 4–5 or 6–8 buffer, was passed through the SPE column at the pre-set flow rate of 0.2 mL min⁻¹. The column effluent was collected. The analyte concentration in the column effluent represents the unrestrained concentration of analyte in the SPE system. The second and final step was the elution of analyte from the SPE system using HCl (1 and 6 M). The analyte concentrations from the sample solution, in the effluent, and in the eluent were measured with GF-AAS. The recovery efficiency was calculated as follows:

\[
\text{Recovery (\%)} = \frac{\text{number of mol of Fe recovered in all fractions}}{\text{number of mol of Fe applied to the column}} \times 100
\]  

(1)

Three replicate measurements per sample were made in all instances. The peak height of the reported signal was proportional to the concentration of the respective iron species and was used for all measurements.
2.4 Instruments

A Hitachi Model Z-8100 GF-AAS (Hitachi, Japan) operating at the 248.3 nm wavelength with a slit width of 0.2 nm and 15.0 mA lamp current was used for analyzing iron concentrations. A temperature program was performed with the following steps: drying at 100 °C, ashing at 1000 °C, atomization at 2700 °C and cleaning at 3000 °C with holding times of 60, 60, 10 and 4 s, respectively. Argon was used as the inert gas at a flow rate of 200 mL min\(^{-1}\) except during the atomization step (30 mL min\(^{-1}\)). After a calibration step, 20 µL of sample and 10 µL of matrix modifier (NiSO\(_4\)) were introduced into the graphite furnace for three replicates of each measurement. The pH of the sample solutions was measured with a Navi F-52 pH meter (Horiba Instruments, Japan) and a combination electrode.

3.0 Results and Discussion

3.1 Comparative study with different SPE materials

The separation efficiencies of Fe(III) from aqueous solutions containing excess ligand with AnaLig TE-01 and different commercially available SPE products were investigated in a non-competitive environment (Fig. 1). Thermodynamic equilibrium calculations suggest that 99.9% of the ambient, dissolved Fe(III) exists as Fe(III) chelates when the ligand concentrations are ~25 times higher in the aqueous system (Bruland and Rue, 2001).

Therefore, a maximal amount of Fe(III) is assumed to be present as the Fe-EDTA complex in the EDTA-spiked sample solutions containing excess EDTA. As shown in Fig. 1, it is apparent that AnaLig TE-01 ensures quantitative extraction of the total amount of Fe(III) from an aqueous solution containing a 100-fold concentration of EDTA spiked with Fe(III). However, the extraction efficiencies of other SPE products are limited to 10–50% for the same sample
solutions. Consequently, AnaLig TE-01 is a more suitable SPE-type system to separate the iron dissolved as the metal-ligand complex in a ligand-rich aqueous environment.

3.2 Effect of pH

The retention of dissolved iron on the AnaLig TE-01 column containing immobilized macrocyclic material was studied as a function of pH. The pH of the aqueous system with Fe(III) and Fe(II) in excess EDTA was maintained in the range of 4–8 with pH 4–5 and 6–8 buffer solutions at 0.1 M concentration. This study was restricted to the pH range 4–8 because EDTA is not very water soluble at very low pH (Ueno et al., 1992), and the increasing solubility of silica gel with pH (Vogelsberger et al., 1992) may dissolve the silica gel base support of AnaLig TE-01 column.

As shown in Fig. 2, quantitative retention of the Fe(III) and Fe(II) complexes from the ligand-rich medium in the pH range 4–8 were at a maximum on the AnaLig TE-01 column. The variation in the pH of the sample solution and the column effluent were insignificant. Therefore, we can conclude that the MRT gel has significant affinity for dissolved iron in a ligand-rich matrix or in iron-ligand complexes, and the iron-ligand complex is stable in the studied pH range. In view of the fact that oceanic water is limited to the pH range of 7 to 9, the majority of the dissolved iron (Fe(III)) is chelated with organic ligands of biological origin (Bruland and Rue, 2001), and further experiments were carried out at pH 8.0 ± 0.2.

3.3 Effect of sample loading flow-rate

The retention of analytes in the SPE system depends upon the flow rate of the metal-fortified sample solution (Bag et al., 1998). The effect of sample loading flow rate on the recovery percentage was examined under optimum conditions. The solution was passed through the SPE
column with the flow rates that were adjusted in the range of 0.2–4.0 mL min⁻¹. As shown in Fig. 3a, retention of Fe(III) on the MRT gel column was quantitative up to a flow rate of 0.25 mL min⁻¹. A gradual decrease in retention with increasing flow rate was observed in the range of 0.3–1.0 mL min⁻¹, and retention decreased to about 60% at higher flow rates (2.0–4.0 mL min⁻¹). This behavior indicates the constant retaining capability of the MRT gel at the initial loading period. Because quantitative, maximum extraction of the analyte is desirable, we applied a flow rate of 0.2 mL min⁻¹ to further experiments to ensure selective pre-concentration of the sample solution for analytical determination.

3.4 Effect of eluent concentration

A satisfactory eluent should effectively elute the extracted analytes using a small volume, which is desired for a high enrichment factor of the analyte, but the eluent should not affect the accurate determination of analytes (Chen et al., 2009). The effect of eluent concentration on the elution of analyte from the MRT gel column was studied by first adding 2 mL of the Fe(III)-spiked, aqueous EDTA solution at pH 8.0 onto the column, and the analyte extracted by the MRT gel column was then eluted using 5 mL of eluent (0.1–6.0 M HCl). The recovery of the analyte was determined with GF-AAS. The Fe(III) recovery percentage was found to increase as the concentration of HCl increased up to a concentration of 0.5 M, at which point the recovery level remained constant (Fig. 3b). This behavior indicates that a HCl concentration equal to 0.5 M is sufficient for quantitative elution of the bound ions in the TE-01 MRT gel column. However, acid concentrations greater than or equal to 5.0 M were recommended as eluent for TE-01 (IBC Advanced Technologies, 2007). Thus, a combination of 1 M HCl (4 mL) and 6 M HCl (1 mL) was selected as eluent for subsequent experiments to ensure quantitative elution of the analyte.
3.5 Effect of metal-ligand stability constants

Ligands form water-soluble metal complexes of high thermodynamic stability (Lim et al., 2005). The high stability of the metal-ligand complexes may influence the extraction performance of the SPE materials. Various investigators have suggested that dissolved iron content in the open oceans exists primarily as Fe-ligand species (Bruland and Rue, 2001), and two classes of Fe(III)-binding natural organic ligands were observed with the following conditional stability constants $K'_{ML}$: $5 \times 10^{12} \text{ M}^{-1}$ ($pK'_{ML} = 12.7$) and $6 \times 10^{11} \text{ M}^{-1}$ ($pK'_{ML} = 11.78$) (Rue and Bruland, 1997).

The effect of the metal-ligand complexes’ $K'_{ML}$ values on the performance of the MRT-SPE system was evaluated for the extraction of Fe(III) from the ligand-rich, Fe(III)-spiked aqueous system. Solutions of ligands in 0.1 M HCl with varying iron complex stability constants ($K_{ML}$), such as NTA, GEDTA, EDTA, DTPA and DFB (Table 2), were added to the iron-spiked solution to prepare each sample solution. The Fe(III)-ligand complex stability at pH 8 was considered during the selection of the chelating ligands. AnaLig TE-01 demonstrated better separation efficiency than the other selected SPE materials (Chelex-100, InterSEP ME-1 and NOBIAS Chelate PA-1) for the metal-ligand complexes of NTA, GEDTA and EDTA (Fig. 4). A significant decrease in the AnaLig TE-01 separation performance was observed for metal-ligand complexes with high $K'_{ML}$ values, e.g., metal-DTPA and metal-DFB. In ocean waters, the $pK'_{ML}$ values for Fe(III)-binding organic ligand classes are limited to a $pK$ of 12.7 (Rue and Bruland, 1997). However, quantitative separation performance for ligands with $pK'_{ML}$ values of up to 22.8 was observed with the proposed MRT-SPE system. Therefore, the thermodynamic stability of the commonly observed Fe(III)-ligand complexes in ocean waters has a negligible effect on the iron separation performance of the AnaLig TE-01.
3.6 Effect of coexisting ions

The interference of other coexisting ions on the separation and preconcentration of dissolved iron was examined under optimal conditions. Various cations, including Na⁺, K⁺, Ca²⁺ and Mg²⁺, and anion species, including Cl⁻, NO₃⁻, CH₃COO⁻, PO₄³⁻, SO₄²⁻ and ClO₄⁻, were added individually to the ligand-rich Fe(III) sample solutions, which were then allowed to equilibrate for 24 h. The studies were carried out in a non-competitive environment by applying 5 mL of the ion-fortified sample at the optimized flow rate with subsequent collection using the appropriate eluent. Variation in pH was negligible between the fortified samples and column effluents. Quantitative recovery of Fe(III) was observed in the presence of the selected cations and anions (Fig. 5). Therefore, it can be concluded that the developed method is reasonably free from interference resulting from coexisting ions that are commonly found in open aqueous systems.

3.7 Extraction capacity

Extraction capacity is an important factor that must be evaluated because it determines how much MRT gel is required for quantitative separation of Fe(III) from a solution. Batch method experiments were used for a capacity study, and the experiments were conducted as follows: 0.5 g of the MRT gel was added to 20 mL of Fe-EDTA solution (Fe(III) – 0.1 mM, ligand: EDTA – 10 mM, matrix: H₂O, pH: 8.0), and the mixture was continuously shaken for one hour. Iron concentrations in the filtrate were determined by GF-AAS. The extraction capacity for Fe(III) under ligand-rich condition was 0.115 ± 0.002 mmol g⁻¹ of AnaLig TE-01 (mean ± SD, n = 3). This result was in good agreement with the certified value 0.1–0.3 mmol g⁻¹ of AnaLig TE-01 (IBC Advanced Technologies, 2007) for the simultaneous complexation of Ag(I), Au(III), Cd(II), Co(II), Cu(II), Fe(II), Hg(II), Ni(II), Pb(II), Pd(II) and Zn(II) from aqueous solutions to concentrate the ions or remove interfering matrices.
3.8 Regeneration and reusability

The regeneration ability and stability of the MRT gel were investigated because regeneration is one of the key factors in evaluating the performance of an extraction material. The experimental results indicated that AnaLig TE-01 is stable during operation of the column, and this material could be regenerated for more than 100 loading and elution cycles without loss of analytical performance. Systems with macrocycles attached onto solid supports (Bradshaw et al., 1988; Izatt, 1997), which is used for the AnaLig TE-01, allow non-destructive separation of metals from the matrix (ligand-rich aqueous solution) and also facilitate the repeated use of the macrocycles because the material can be regenerated after each use. Thus, the initial high cost of MRT gel synthesis is amortized over time.

3.9 Analytical characteristics

GF-AAS was used to measure the iron concentrations in the MRT gel treated, fortified aqueous samples. Under optimum conditions, the linear range was found to be 0.01–0.11 µg mL\(^{-1}\). The method detection limits, as calculated from three times the standard deviation \((n = 15)\) of the blank, was 0.02 µg L\(^{-1}\). Based on the relative standard deviation, the precision of the method was calculated to be 1.4% from 10 replicate measurements at a concentration of 0.4 µM Fe(III).

3.10 Recovery test with artificial seawater

Artificial seawater was prepared according to Lyman and Fleming (1940) and used to evaluate the ion-recovery performance of the MRT-SPE column in the presence of multiple interfering ions. Artificial seawater samples spiked with 0.1 mM Fe(III) and 10 mM EDTA were passed through the MRT-SPE system at pH 8. The recovery of Fe(III) was observed to be 96 ± 3% \((n = 3)\).
4.0 Conclusions

MRT gel, a column-packed, immobilized macrocyclic material, was used for the separation/preconcentration of dissolved iron from an aqueous system containing excess ligand, and GF-AAS was used to determine the concentrations of eluted iron. Quantitative collection of the dissolved iron was obtained under the following optimized conditions: pH range: 4–8; flow rate: 0.2 mL min$^{-1}$; eluent: 1 M and 6 M HCl.

Compared with previously tested SPE materials, MRT gel possesses several advantages. MRT gel is chemically stable with excellent separation properties and extraction capacity. It can be used repeatedly with little loss of performance. Separation of the dissolved iron is rapid, reproducible, and efficient. In addition, the presence of large concentrations of other coexisting ions had no affect on the separation process.

In natural oceanic systems, dissolved Fe(III) primarily exists as organically-bound Fe-ligand species, and the total organic ligand concentrations are ~25 times higher than total iron concentrations. In freshwater systems, Fe(III) often forms complexes with humic substances. Conventional SPE systems may not be applicable for the quantitative determination of dissolved iron in natural bodies of water due to competition with strong Fe(III)-binding organic ligands that limits their accuracy. This limitation is minimized with the proposed MRT-gel separation technique, and it is expected that the total dissolved iron content can be accurately measured with this technique in the presence of an excess of strong Fe(III)-binding organic ligands. Importantly, the quantitative separation performance of the proposed MRT-SPE system was achieved for metal-ligand complexes with a maximum $pK'_{ML}$ of 22.8. Current work is underway to determine ultra-trace amounts of iron in an aqueous system containing excess organic ligands using the MRT gels; the goal of this work is to develop a technique for practical
separation/preconcentration and subsequent determination of dissolved iron in oceanic water samples.

Acknowledgement

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Table 1: Different SPE materials used in this study
<table>
<thead>
<tr>
<th>SPE material type</th>
<th>Commercial Name</th>
<th>Base Support</th>
<th>Functional group</th>
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<tbody>
<tr>
<td>MRT Gel</td>
<td>AnaLig TE-01</td>
<td>Silica gel</td>
<td>Crown ether</td>
</tr>
<tr>
<td>Ion-exchange resin</td>
<td>NOBIAS Ion SC-1</td>
<td>Hydrophilic methacrylate</td>
<td>Sulfonic acid</td>
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<tr>
<td></td>
<td>NOBIAS Ion SA-1</td>
<td>Hydrophilic methacrylate</td>
<td>Quaternized amine</td>
</tr>
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<td>Chelate resin</td>
<td>Chelex-100</td>
<td>Styrene divinylbenzene</td>
<td>Iminodiacetic acid</td>
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<td>MetaSEP ME-1</td>
<td>Methacrylate polymer</td>
<td>Iminodiacetic acid</td>
</tr>
<tr>
<td></td>
<td>MetaSEP ME-2</td>
<td>Methacrylate polymer</td>
<td>Iminodiacetic acid +a</td>
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<td></td>
<td>MetaSEP ME-3</td>
<td>Methacrylate polymer</td>
<td>Iminodiacetic acid +b</td>
</tr>
<tr>
<td></td>
<td>NOBIAS Chelate PA-1</td>
<td>Hydrophilic methacrylate</td>
<td>Polyamino-polycarboxylic acid</td>
</tr>
<tr>
<td></td>
<td>NOBIAS Chelate PB-1</td>
<td>Divinylbenzene/methacrylate polymer</td>
<td>Polyamino-polycarboxylic acid</td>
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</tbody>
</table>

Table 2: Acid dissociation constants ($pK_a$), stability constants ($K_{ML}$) and conditional stability constants ($K'_{ML}$) of Fe(III)-ligand complexes at 25 °C
<table>
<thead>
<tr>
<th>Ligand</th>
<th>$K_{\text{ML}}$</th>
<th>$pK_a$</th>
<th>$K'_{\text{ML}}$ (at pH 8)</th>
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<tbody>
<tr>
<td>NTA</td>
<td>15.9&lt;sup&gt;a&lt;/sup&gt;</td>
<td>1.89, 2.49, 9.73</td>
<td>14.1</td>
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<tr>
<td>GEDTA</td>
<td>20.5&lt;sup&gt;a&lt;/sup&gt;</td>
<td>2.00, 2.68, 8.85, 9.46</td>
<td>18.1</td>
</tr>
<tr>
<td>EDTA</td>
<td>25.0&lt;sup&gt;a&lt;/sup&gt;</td>
<td>1.99, 2.67, 6.16, 10.26</td>
<td>22.8</td>
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<tr>
<td>DTPA</td>
<td>28.6&lt;sup&gt;b&lt;/sup&gt;</td>
<td>2.14, 2.38, 4.26, 8.60, 10.53</td>
<td>25.4</td>
</tr>
<tr>
<td>DFB</td>
<td>31.9&lt;sup&gt;c&lt;/sup&gt;</td>
<td>8.39, 9.03, 9.70</td>
<td>29.0</td>
</tr>
</tbody>
</table>

<sup>a</sup>Martell and Smith (1974), <sup>b</sup>Sillen and Martell (1964; 1993), <sup>c</sup>Morel and Hering (1993)
Figure 1: Comparative performance of AnaLig TE-01 with different commercial SPE materials for aqueous metal solution containing excess ligand \( (n = 3) \). Sample solution: Fe(III) – 0.1 mM, ligand: EDTA – 10 mM, matrix: H\(_2\)O, pH: 8.0, sample volume: 5 mL, flow rate: 0.2 mL min\(^{-1}\), eluent: 1 M HCl (4 mL) + 6 M HCl (1 mL). ‘Sample’ denotes the analyte concentration in the column effluent collected after passing the sample solution through the MRT-SPE system; ‘elute’ denotes the analyte concentration in the collected eluent.
Figure 2: Effect of pH on the recovery percentage ($n = 3$). Sample solution: (a) Fe(III) – 0.1 mM and (b) Fe(II) – 0.1 mM, ligand: EDTA – 10 mM, matrix: H$_2$O, pH: 4.0–8.0, sample volume: 5 mL, flow rate: 0.2 mL min$^{-1}$, eluent: 1 M HCl (4 mL) + 6 M HCl (1 mL). ‘Sample’ denotes the analyte concentration in the column effluent collected after passing the sample solution through the MRT-SPE system; ‘elute’ denotes the analyte concentration in the collected eluent.
Figure 3: Effect of (a) sample loading flow rate and (b) eluent concentration on the recovery percentage of analyte ($n = 3$). Sample solution: Fe(III) – 0.1 mM, ligand: EDTA – 10 mM, matrix: H$_2$O, pH: 8.0, sample volume: 5 mL, flow rate: (a) 0.2–4.0 mL min$^{-1}$ and (b) 0.2 mL min$^{-1}$, eluent: (a) 1 M HCl (4 mL) + 6 M HCl (1 mL) and (b) HCl (0.1–6.0 M) – 5 mL.
Figure 4: Effect of metal-ligand stability constants on the performance of SPE materials (n = 3).
Sample solution: Fe(III) – 0.1 mM, ligand: NTA, GEDTA, EDTA, DTPA, DFB – 10 mM, matrix: H2O, pH: 8.0, sample volume: 5 mL, flow rate: 0.2 mL min⁻¹, eluent: 1 M HCl (4 mL) + 6 M HCl (1 mL).
Figure 5: Effect of coexisting ions on the performance of AnaLig TE-01 ($n = 3$). Sample solution: Fe(III) – 0.1 mM, ligand: EDTA – 10 mM, ion concentration – 10 mM, matrix: H$_2$O, pH: 8.0, sample volume: 5 mL, flow rate: 0.2 mL min$^{-1}$, eluent: 1 M HCl (4 mL) + 6 M HCl (1 mL). ‘Sample’ denotes the analyte concentration in the column effluent collected after passing the sample solution through the MRT-SPE system; ‘elute’ denotes the analyte concentration in the collected eluent.